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Functional brain networks underlying latent inhibition of conditioned disgust in rats

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Abstract

The present experiment examined the neuronal networks involved in the latent inhibition of conditioned disgust by measuring brain oxidative metabolism. Rats were given nonreinforced intraoral (IO) exposure to saccharin (exposed groups) or water (non-exposed groups) followed by a conditioning trial in which the animals received an infusion of saccharin paired (or unpaired) with LiCl. On testing, taste reactivity responses displayed by the rats during the infusion of the saccharin were examined. Behavioral data showed that preexposure to saccharin attenuated the development of LiCl-induced conditioned disgust reactions, indicating that the effects of taste aversion on hedonic taste reactivity had been reduced. With respect to cumulative oxidative metabolic activity across the whole study period, the parabrachial nucleus was the only single region examined which showed differential activity between groups which received saccharin-LiCl pairings with and without prior non-reinforced saccharin exposure, suggesting a key role in the effects of latent inhibition of taste aversion learning. In addition, many functional connections between brain regions were revealed through correlational analysis of metabolic activity, in particular an accumbens-amygdala interaction that may be involved in both positive and negative hedonic responses.

Keywords: taste aversion, conditioned disgust, taste reactivity, latent inhibition, cytochrome oxidase.

Abbreviations

Adenosine triphosphate (ATP)

Amygdala, basolateral nucleus (BLA)

Amygdala, central nucleus (CeA)

Bed nucleus of stria terminalis (BNST)

Cingulate cortex (Cg)

Conditioned stimulus (CS)

Cytochrome c oxidase (CO)

Infralimbic cortex (IL)

Insular cortex (IC)

Intraoral (IO)

Lithium Chloride (LiCl)

Medial prefrontal cortex (mPFC)

Nucleus Accumbens (NAcb)

Parabrachial nucleus (PBN)

Prelimbic cortex (PrL)

Sodium Chloride (NaCl)

Taste reactivity (TR)

Unconditioned stimulus (US)

Ventral tegmental area (VTA)

Ventralis posterolateral thalamic nucleus (VPL)

Ventralis posteromedial thalamic nucleus (VPM)

1. Introduction

Nausea and vomiting resulting from chemotherapy treatments for cancer is reported as the most distressing side-effect that can cause patients to discontinue treatment [1,2]. In addition to the effects directly induced by the drug-induced nausea, patients often develop anticipatory nausea and vomiting to contextual stimuli related to the hospital environment, and effect that can be explained in terms of classical conditioning [3,4]. The cues that constitute the clinic (smell, sounds, and even the staff) can become associated with the nausea-inducing treatment and, as a consequence of the formation of such association, these stimuli may become able to evoke a response (the conditioned response, CR) that is nausea and vomiting [5]. Theoretical analysis of the conditioning processes involved in the acquisition of context aversions in animals suggests possible behavioral strategies that might be used in the alleviation of anticipatory nausea and vomiting [6,7]. In particular, one effective way to attenuate anticipatory nausea is preventing the formation of context-nausea association by giving extensive prior exposure to the contextual cues, a conditioning phenomenon referred as the latent inhibition effect [8,9].

The initial goal of the current study was to examine the brain regions involved in latent inhibition of nausea-induced conditioned disgust. In the taste aversion paradigm, the latent inhibition effect is defined as a retardation of conditioning due to extensive prior exposure to the flavor that is to be paired with a drug with emetic properties (e.g., lithium chloride, LiCl). Therefore, the contribution of flavor novelty/familiarity in the development of conditioned disgust can be examined with this paradigm. Whilst the latent inhibition effect has been extensively evaluated using voluntary fluid ingestion, the current study aims to evaluate this effect by measuring conditioned disgust reactions elicited by a nausea-paired flavor. Flavors

can be administered directly into the rat's mouth through a cannula implanted in the oral cavity. This procedure allows the orofacial reactions displayed by the rats during the intraoral infusion of the fluid to be recorded and analyzed, a method termed the taste reactivity (TR) test [10]. The TR method measures changes in the hedonic value of a flavor due to aversive visceral experience. In particular, rodents display conditioned gaping reactions (a disgust reaction selective to nausea) during re-exposure to the conditioned flavor. TR is a particularly suitable procedure for measuring conditioned taste aversion because it is selective to disgust, in contrast to decreases in the voluntary intake of the flavor which may simply reflect taste avoidance without a change in hedonic responses [11,12,13]. In recent studies of our laboratory using taste reactivity and licking analysis methods we found that flavor preexposure attenuates the effects of taste aversion learning on both flavor consumption and taste palatability [14]. Based upon these findings, in the present study we explored the neural networks involved in latent inhibition of conditioned disgust by measuring changes in metabolic activity through cytochrome c oxidative (CO) histochemistry in brain regions generally accepted as being involved in conditioned taste aversion. These brain regions may be divided into two systems, one related to the processing of taste qualities and the other to the reward system. The taste recognition system includes the nucleus of the solitary tract, parabrachial nucleus, the ventralis posteromedial thalamic nucleus, and the insular cortex. The reward systems would include mainly the ventral tegmental area and the accumbens. Prefrontal cortex and amygdala are the candidates to be the interface regions between these two systems.[15,16,17] Despite the evidence that flavor familiarity influences neural activity in conditioned taste aversion as assessed by consumption tests, it is unclear what brain regions are activated in response to hedonic shifts in this learning paradigm. In addition to the fact that latent inhibition is of interest in its own right, the fact that pre-training exposure to the taste stimulus retards conditioning affords a direct comparison of the brain effects of

1 identical taste-nausea pairings between animals that form strong aversions (i.e. the non-
2 exposed group) with those that will not form a strong aversion (i.e. the exposed group).
3

4 The neuronal underpinnings of conditioned taste aversion have been widely examined
5 using c-fos immunocytochemistry [18,19,20]. However, the special functional properties of
6 taste aversion learning may require alternative methods to fully explore the physiological
7 bases of this learning. In contrast to other forms of learning, taste aversions are well known to
8 form in a single trial and can be formed despite long-delays between the flavor and the illness
9 [21, 22]. Therefore, a method that assess long term changes in the brain activity would be
10 particularly valuable. Due to the close relationship between the electrical activity of neurons
11 and oxidative energy metabolism, an alternative way to study brain activity is by using
12 histochemical techniques such as the mitochondrial enzyme cytochrome c oxidase (CO)
13 [23,24]. Cytochrome c oxidase is a mitochondrial enzyme implicated in the oxidative
14 phosphorylation process in which adenosine triphosphate (ATP) is generated. This
15 histochemical technique identifies structures involved in long-term metabolic changes such as
16 increased levels of enzymes or neurotransmitters, membrane protein synthesis, morphological
17 changes (high postsynaptic density) and neuronal electrical increased activity, reflecting other
18 process that also require ATP in the cell. CO histochemistry has been used in studies of
19 learning and memory in a variety of animal species [25,26,27,28,29]. The use of CO
20 histochemistry provides a range of advantages over the use of early gene expression as c-Fos
21 immunohistochemistry, in particular c-Fos methodology indicates acute (seconds to minutes)
22 transient changes in neural activation evoked by a stimulus as a result of a particular
23 intervention and thus might not fully capture the processes involved conditioned taste
24 aversion where conditioning can occur with long delays between flavor exposure and the
25 induction of illness. In addition, CO histochemistry allows the assessment of cumulative
26 changes throughout the learning process, which is critical in understanding the processes
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underpinning latent inhibition, where non-reinforced initial exposure can occur a matter of days before conditioning and testing. CO activity provides an accurate assessment of the metabolic history of brain regions because sustained increased or decreased in neuronal activity lead to corresponding changes in CO activity [30,31,32]. As such, it reflects the temporal dynamics of brain metabolism over the acquisition and retrieval of learning, not only in specific brain regions but also in terms of their functional connectivity [33].

Therefore, the goal of the present study was to examine, using CO histochemistry, changes in brain metabolic activity following latent inhibition of conditioned disgust. In particular, because CO activity is sensitive to manipulations between the day of measurement and up to a week before [27], this will reflect the impact of the whole behavioral preexposure, conditioning, and test procedure performed here. More specifically, we attempt to determine the functional connectivity between different brain regions involved in the development of nausea-induced conditioned disgust.

2. Materials and methods

2.1. Subjects

The subjects were 31 male Wistar rats weighing 190-339 g (258 g mean weight) at the start of the experiment. Upon arrival, they were individually housed in opaque plastic cages in a room maintained at 21° C with a 12/12h light-dark cycle. All experimental manipulations were performed during the light portion of the cycle. Food and water available in the home cages throughout the experimental procedures. All experimental procedures were in accordance with guidelines for the care and use of laboratory animal of the Spanish (RD 12301/2005) and European (86/609/EEC) regulations concerning animal experimentation.

2.2. Cannulation surgery

In order to implant intraoral cannula, the rats were anesthetized with ketamine (50 mg/kg) combined with metedomidine hydrochloride (0, 15 mg/kg). A 3-cm square of fur was shaved on the back of the neck and the area was then wiped with an antiseptic product (Betadine). A thin-walled 15-gauge stainless steel needle was inserted at the back of the neck, routed underneath the skin, and brought out inside the mouth behind the first molar. The cannula was inserted through the needle, which was then removed. Cannulae consisted of intramedic polyethylene tubing with an inner diameter of 0.86 mm and an outer diameter of 1.27 mm. To prevent the cannula from moving out of place, two elastic discs were placed over the tubing and drawn to the exposed skin at the back of the neck for the purpose of stabilizing the cannula. After the cannulation surgery, the rats were monitored for three days. The cannulas are cleaned daily with distilled water and the rats were injected subcutaneously with an antibiotic (enrofloxacin 0.3 mg/kg) and anti-inflammatory (ketoprofen 1.5 mg/kg).

2.3. Apparatus

For the purpose of fluid infusion, the cannula was connected to an infusion pump (Kd Scientific Inc., MA) by slipping the tubing of the cannula inside a second polyethylene tubing (inner diameter 1.19 mm; outer diameter 1.79 mm) attached to the infusion pump. Intraoral infusions were carried out in a conditioning chamber that was situated in a room without natural light. The chamber consisted of a wooden frame painted black (150 x 64 x 50 cm) with two points of light (50w) one on each side of the chamber. The top of the apparatus consisted of a sheet of transparent plastic (Plexiglas) and a mirror beneath the chamber (60 x 45 cm) inclined at 45 degrees to facilitate viewing of the ventral surface of the rat during fluid infusion. The rats were placed in the top of the conditioning apparatus within a square compartment (22.5 x 26 x 20 cm) made of Plexiglas. This compartment had a lid of dark Plexiglas with a central hole through which we introduced the plastic tube connected to the

infusion pump. The other end of the plastic tube was connected to the oral cannula during the IO infusion. The sessions were recorded with a video camera (Sony Handicam) connected to a computer that enabled recordings to be directly stored (see Grill [34] for a schematic representation of the apparatus for videotaping taste reactivity).

Orofacial reactions made by the animals during the infusion of fluids were recorded manually quantified and scored by two raters with the software The Observer XT 9.0 (Noldus Information Technology). The behaviors scored included the frequency of the disgust reactions of gaping (rapid, large-amplitude opening of the mandible with retraction of the corners of the mouth), chin rubbing (mouth or chin in direct contact with the floor or wall of the chamber and body projected forward), and paw treading (forward and backward movement of the forepaws in synchronous alternation). These scores were summed to provide a total disgust reaction score. The ingestion reactions of tongue protrusions (extension of the tongue out the mouth) and mouth movements (movement of the lower mandible without opening the mouth) were also scored. The number of seconds that the rats displayed the reactions of tongue protrusions and mouth movements was used as the appetitive reaction score.

2.4. Behavioral procedure

After recovery from the cannula implantation surgery, rats were randomly assigned to four groups: preexposed LiCl-paired (PE-CTA, n=8), non-preexposed LiCl-paired (NPE-CTA, n=8), preexposed LiCl-explicitly unpaired (PE-CONT, n=7) and non-preexposed LiCl-explicitly unpaired (NPE-CONT, n=8). Experimental sessions were initiated at 10:00 AM each day. Two days after the cannulation surgery the rats were habituated to the infusion procedure, receiving a 5 ml water infusion (rate of infusion 1 ml/min) in the conditioning apparatus and returned to their home cages. The next 4 days constituted the flavor preexposure

phase (see Table 1). During these sessions, the rats in groups PE-CTA and PE-CONT were intraorally infused with 0.1% saccharin for 5 min at a rate of 1 ml/min whereas those in groups NPE-CTA and NPE-CONT received water infusions. On the next day, the conditioning trial was carried out. This trial consisted of a single session of an IO saccharin infusion (0.1%) for 5 min at a rate of 1 ml/min. Immediately following the saccharin infusion, rats from groups PE-CTA and NPE-CTA were injected with LiCl (20 ml/kg, 0.15 M) whereas groups PE-CONT and NPE-CONT received the LiCl injection 24-h after the saccharin infusion. Subsequently, the rats were returned to their home cages in order to recover from the LiCl effects. After a recovery day in the home cages, the TR test was carried out. This test consisted of an intraoral administration of 5 ml of saccharin (0.1%) at a rate of 1 ml/min in the conditioning chamber while the rat's orofacial reactions were videotaped.

TABLE 1

2.5. Cytochrome oxidase histochemistry

The changes in metabolic activity were analyzed using CO histochemistry. The procedure followed was described by Wong-Riley [23] based on certain modifications proposed by Gonzalez-Lima and Jones [24]. The structures were selected for analysis using the atlas of Paxinos and Watson [35]. Ninety minutes after the taste reactivity test, the rats were sacrificed by decapitation. Their brains were rapidly removed and frozen by immersion in isopentane (Sigma-Aldrich, Spain) at -70° C for two minutes and stored at -40° C to prevent tissue damage with loss of enzyme activity.

Subsequently, the brains were sectioned coronally with a microtome cryostat (Microm International GmbH, HM 505E, Heidelberg, Germany) in sections (30 µm thick) that were placed on slides previously cleaned with alcohol at 100%. The sections were fixed with 0.5 % (v/v) glutaraldehyde and 10% (W/V) sucrose in 0.1 M phosphate buffer, pH 7.6. After that,

1 sections were rinsed in 0.1 M phosphate buffer with 10% (W/V) sucrose. Following this
2 procedure, they were immersed in 0.05 M Tris buffered solution, pH 7.6 containing 275 mg/l
3 cobalt chloride, 10% (w/v) sucrose and 0.5 % (v/v) dymethylsulfoxide. The sections were
4 incubated in darkness for 2 h at 37° C in a staining bath containing 0.06 g cytochrome c,
5 0.016 g catalase, 40 g sucrose, 2 ml dimethylsulfoxide and 0.4 g diaminobenzidine
6 tetrahydrochloride (Sigma-Aldrich, Spain) dissolved in 800 ml of phosphate buffer (pH 7.6;
7 0.1 M). Finally, the sections were fixed at 4% (v/v) buffered formalin with 10% (w/v)
8 sucrose, dehydrated in alcohol chain and coverslipped with Entellan (Merck, Germany). A
9 series of sections of rat brain cut at different thicknesses (10, 30, 50 y 70 µm) were included
10 together with brain tissue in each bath. These sections were used as standards to control for
11 staining variability across different incubation baths, and were obtained from rat brain
12 homogenates. The tissue homogenates were obtained from the brains of 12 adult male Wistar
13 rats killed by decapitation and homogenized on ice. To establish comparisons in the staining
14 of sections from different staining baths, measurements were taken from CO stained brain
15 homogenate standards. These activity values allowed for the construction of a linear
16 regression curve to convert measured values of optical density of the selected structures in
17 cytochrome oxidase activity units (1 unit: 1 µmol of cytochrome c oxidase/min/g tissue wet
18 weight at 23° C), and were calculated for each incubation bath.

19 Using an image processing system (Leica Q550, Germany) with a light source
20 (LeicaDM-RHC) connected to a camera CCD (Lohu, Japan) and image analysis software
21 (Leica Q-Win), relative optical density (OD) readings were obtained from the brain regions
22 of interest: medial prefrontal cortex (mPFC) including cingulate (Cg), prelimbic (PrL) and
23 infralimbic cortex (ILc); insular cortex (IC); bed nucleus of stria terminalis (BNST); central
24 and basolateral amygdala (CeA and BLA); ventral tegmental area (VTA); parabrachial
25 nucleus (PBN); ventralis posteromedial thalamic nucleus (VPM) and core and shell nucleus

of accumbens (NAcb). Figure 1 represent microphotographs showing examples of coronal CO stained sections and schematic representations of the selected brain regions. A total of 25 measurements were taken per region. The relative optical density was measured by taking 5 non-overlapping regions in each section for 5 consecutive sections by using a square-shaped sampling window previously adjusted for each region size (see example of PrL sampling on Figure 1). Then, measures were averaged to obtain one data point per region for each animal (group data shown in Table 2).

FIGURE 1

2.6. Statistical analysis

The data were analyzed using SPSS version 19.0 (SPSS Inc., Chicago, USA). Data for behavior scored in taste reactivity test were analyzed by a 2 (preexposure) x 2 (conditioning) analysis of variance (ANOVA). In all cases, a significance criterion of $p < 0.05$ was used.

The inter-rater reliability ($r's > 0.87$) for each behavior scored was highly significant.

Differences in CO staining of each brain region between the selected groups were analyzed by a similar analysis of variance (ANOVA). In order to examine functional connectivity

between brain regions, pairwise correlations in CO activity were analyzed by calculating

Pearson's product moment correlation coefficients ($p < 0.05$) with CO activity normalized values (for each animal, CO activity of each region divided by mean activity in all regions).

A "Jackknife" procedure [36] was used such that pairwise correlations were first calculated including all animals in the analysis. Then, one subject was removed each time and correlations were calculated anew. Only correlations present in all comparisons made without one of the subjects (and all subjects) are reported as significant. This procedure minimizes the risk of spurious correlations being produced by outliers in the data.

3. Results

3.1. Taste reactivity test

Figure 2 (top panel) presents the mean number of conditioned disgust reactions displayed by rats during the 5-min intraoral infusion of saccharin in the TR test. It can be seen that rats in Group NPE-CTA showed a higher rate of aversive reactions than animals in the remaining groups, indicating that non-reinforced exposure to saccharin attenuated the establishment of LiCl-induced conditioned disgust. The two-way ANOVA conducted with these scores revealed significant main effects of preexposure [$F(1,27) = 8.44$; $p = 0.007$], conditioning [$F(1,27) = 23.05$; $p < 0.001$], and a significant preexposure x conditioning interaction [$F(1,27) = 7.74$; $p = 0.010$]. Simple effects analysis revealed that Group NPE-CTA displayed a higher rate of aversive reactions to the saccharin infusion than Group PE-CTA [$F(1,27) = 16.75$; $p < 0.001$], and that non-conditioned groups did not differ from one another ($F < 1$). These results confirm that repeated exposure to saccharin solution retards the development of conditioned nausea to that flavor. Figure 2 (bottom panel) shows the mean duration of appetitive reactions displayed by the rats during the TR test. The two-way ANOVA conducted with these data revealed a main effect of preexposure [$F(1,27) = 15.32$; $p = 0.001$] and conditioning [$F(1,27) = 13.27$; $p = 0.001$], and a significant preexposure x conditioning interaction [$F(1,27) = 7.79$; $p = 0.010$]. Simple effects analysis revealed that Group NPE-CTA displayed a lower rate of appetitive reactions to the saccharin infusion than Group PE-CTA [$F(1,27) = 23.29$; $p < 0.001$], and non-conditioned groups did not differ from one another ($F < 1$). These results show that prior exposure to the to-be conditioned flavor not only attenuates the development of conditioned disgust, but also protects against the reduction in positive hedonic reactions.

TABLE 2 & FIGURE 2

3.2. CO activity

Table 2 shows the CO activity measured in the brain areas of interest. The 2 (preexposure) x 2 (conditioning) ANOVA conducted on these data revealed a significant main effect of conditioning in mPFC subregions [cingulate, prelimbic and infralimbic; $F_s(1,27) > 4.59$; $p_s < 0.041$], in bed nucleus of stria terminalis [$F(1,27) = 12.55$; $p < 0.001$], basolateral amygdala [$F(1,27) = 4.28$; $p = 0.048$], ventral tegmental area [$F(1,27) = 11.064$; $p = 0.003$] and PBN [$F(1,27) = 4.07$; $p = 0.05$], but no main effect of preexposure or preexposure x conditioning interaction ($F_s < 1$), with the exception of the PBN region which had a significant preexposure x conditioning interaction [$F(1,27) = 5.06$; $p = 0.033$]. Simple main effects analysis revealed lower CO activity in PBN for the latent inhibition group; that is, Group PE-CTA [$F(1,27) = 4.85$; $p = 0.036$] compared to NPE-CTA group. There was also no differences between Group PE-CONT and Group NPE-CONT [$F(1,27) = 1.01$; $p = 0.326$]. This result shows that a conditioned change in the hedonic value of a flavor is related with long term changes in PBN activity, while the pairing of the flavor with illness resulted in lower activity in a number of brain regions irrespective of whether this resulted in a hedonic shift.

Interregional correlations of CO activity were analyzed in order to establish different brain networks in each group. A network is defined by a set of structures linked by correlations and independent of correlational links with other structures. Figure 3 shows significant cross-correlations of CO activity for the different groups (r values). These correlations revealed two brain networks for Group PE-CONT. The first one included the PBN and ventral tegmental area with a positive correlation. The second network comprised a prefrontal cortical region, the cingulate cortices, which showed a positive correlation with the

nucleus accumbens core. In turn, NAcb shell had a positive correlation with CeA. This latter area also had a negative correlation with the ventral posterolateral thalamus, which showed a negative correlation with the prefrontal cortex and NAcb shell.

FIGURE 3

For the NPE-CONT group, correlations also revealed two brain networks. First, there was a positive correlation between the basolateral amygdala and PBN region. Subsequent correlations show a brain network that comprised cingulate cortex, which had a positive correlation with the thalamus and with the NAcb core and shell. NAcb shell also showed a negative correlation with the ventral tegmental area.

Groups which experienced saccharin-lithium pairings (PE-CTA and NPE-CTA) showed few positive correlations between brain regions. In particular, the PE group showed positive correlations between prelimbic and infralimbic, NAcb core and thalamus, and finally between the PBN and tegmental areas. NPE-CTA group had positive correlations between the prefrontal areas (Cg, PrL and Ilc), NAcb shell and central amygdala, and between PBN and tegmental areas.

4. Discussion

The aim of the present study was to examine the functional brain networks involved in nausea-induced conditioned disgust reactions. For this purpose, a latent inhibition paradigm was used to measure, at both the behavioral and neuronal level, the effect of non-reinforced flavor exposure on the subsequent establishment of a conditioned taste aversion. Behavioral data revealed that non-reinforced exposure to a saccharin solution prior to its pairing with nausea attenuates the development of conditioned disgust reactions to the flavor. This effect

1 is consistent with previous latent inhibition studies using consumption tests [8,9] and from
2 our own laboratory using TR [14] or licking microstructure [37] procedures.
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4 With respect to overall differences in regional CO activity, there were reductions in
5 brain CO activity in BLA, BNST, PBN, VTA and prefrontal areas in groups which
6 experienced saccharin-lithium pairings (PE-CTA and NPE-CTA) regardless of whether these
7 pairings resulted in a change in the hedonic reaction to the taste. The only brain region
8 displaying changes in activity relating to the change in taste reactivity was the PBN. Because
9 the taste reactivity change was determined by the presence or absence of latent inhibition, this
10 result highlights the role of the PBN as a key mediator of the way that latent inhibition in
11 attenuates or prevents CS-US associations forming in conditioned taste aversion.
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24 Turning to the analysis of the connectivity between brain regions, our data shows a
25 positive correlation between the PBN and VTA in both conditioned groups, that is, groups
26 PE-CTA and NPE-CTA, regardless of whether latent inhibition prevented the development of
27 conditioned disgust reactions. It should be noted that groups PE-CTA and NPE-CTA showed
28 decreased CO activity in the PBN and VTA compared to the remaining groups. As already
29 indicated, PBN is the candidate structure for CS-US associations, whereas VTA is involved
30 in fluid reward (regardless of negative or positive hedonic fluid value) [38]. In addition, the
31 data show a positive correlation between the mPFC subregions for the novel CS-US
32 association, and a PrL- Ilc subregions correlation in familiar CS-US associations. This may
33 indicate that conditioned taste aversion learning requires the involvement of all subregions of
34 the mPFC for taste memory formation. This is consistent with other lesion studies in which
35 an impairment of conditioned taste aversion is observed as a result of mPFC injury [39, 40].
36 In addition, we found a negative correlation between NAc and thalamus nuclei in group PE-
37 CTA, indicating that both structures may mediate the hedonic shift from positive to negative,
38 and the data implicate the involvement of NAc in this shift [22]. Perhaps more interesting is
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1 the amygdala-NAcb interaction found in Group NPE-CTA – the only group to show a
2 behavioral taste aversion. It is known that both NAcb and amygdala regions are essential for
3 expression of conditioned taste aversion [41]. Some studies have associated the amygdala
4 with the processing of visceral information, given that LiCl injections increase c-fos activity
5 in this region [38], linking its function to the recognition of US information. However, other
6 studies suggest a central role for this structure in the formation of CS-US association, relating
7 BLA with aversive conditioning [42] and with the formation of fear learning in general. But
8 CeA is the main output route for other brain regions related to the expression of specific
9 emotional responses [43], and its specific role remains unclear. Our data may link this region
10 to the activity of the NAcb in the formation of taste aversion memory. However, since a
11 similar relation was found in the PE-CONT group, amygdala-NAcb connections may be
12 involved in both appetitive and aversive rewards, given that this interaction has been found in
13 the group for which the saccharin was highly appetitive (PE-CONT) or highly aversive
14 (NPE-CTA).

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34 Considering group PE-CONT, which had repeated experience with a pleasant taste, in
35 more detail, the analysis revealed two main brain networks (see Figure 3). First, a positive
36 correlation was shown between the PBN and VTA. This is consistent with the suggestion that
37 palatable fluids such as saccharin or sucrose activate VTA neurons [44, 45] and its lesions
38 selectively reduce consumption of a preferred sucrose solution [46]. This relationship may be
39 due to the PBN axons that pass through the VTA [47] and the fact that this area is also
40 connected with other limbic structures [48]. Indeed, the PBN might influence limbic
41 structures such as the NAcb via direct connections with the VTA [49].

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53 The second network comprised the NAcb (shell and core), CeA and Cg interactions, all
54 of which showed negative correlations with the thalamus nuclei. Several studies have shown
55 that NAcb receives afferent inputs from mPFC and amygdala structures (see for instance,
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[41]). Indeed, long-lasting increase in dopamine release in NAcB is indirectly modulated by amygdala-induced dopamine responses in mPFC [50]. NAcB has been associated with reinforced learning, i.e. an association between pleasant flavors (or unpleasant) and its consequences. In particular, NAcB activity increased with intraoral infusions of pleasant flavors [51, 52]. Our data suggest that brain regions involved in reward (NAcB), regions that send information of sensorial properties of flavors (thalamus) to the main candidates for integrate reward and sensory information (amygdala and mPFC) [22], were functionally paired when a familiar and pleasant flavor is intraorally administered.

Brain networks involved in the processing of familiar pleasant solutions differ from those involved in processing novel solutions. For group NPE-CONT, two brain networks may be established, a PBN-amygdala interaction, and a second network that comprises the VTA, NAcB, Cg and thalamus. These results are compatible with other research studies [50] in which the neural pathway of the brain reward system is described as being mainly comprised of the VTA and NAcB. As previously mentioned NAcB is interconnected with PFC which is associated with feeding control and receives afferents from the thalamus via IC. Thalamus (VPM) on the other hand, is related to the processing of novel flavors, given that its lesions disrupt neophobia [22]. Taken together, our results may suggest that information from the reward pathway (VTA and NAcB) and sensorial information (thalamus) regarding flavor novelty may converge in the PFC to be integrated for the formation of flavor memories. Additionally, we have found a positive correlation between PBN and BLA. Previous studies revealed that exposure to a novel, but not familiar, saccharin solution induced robust increases of c-fos activity in the PBN and CeA but not in the BLA [53]. However, the BLA may be involved in neophobia [54], and therefore in the processing of flavor novelty, given that injuries in this area disrupts CS-US associations with a novel, but not familiar CS ([55] for a review). It is also accepted that initial integration of gustatory and visceral signals takes

place in PBN [56, 57, 41] and as well as the amygdala, PBN plays a crucial role in the processing of novel flavors and the consequences derived from their intake [58, 59, 60]. This hypothesis is supported by our data, in which the PBN reduce its activity as a result of the latent inhibition of the conditioned flavor.

It is important to emphasize that the CO histochemistry technique assesses changes throughout the learning procedure. This means that in contrast to other techniques (early gene expression, for instance) in which activity changes are measured at a specified point in time, the CO technique allows for the measurement of accumulative changes. This may result in unchanged activity in brain regions traditionally related to latent inhibition in conditioned taste aversion, or alternatively, these brain regions may not elicit long-term changes in CO activity.

5. Conclusion

The present study showed that non-reinforced exposure to saccharin prior to the acquisition of a LiCl-induced saccharin aversion attenuates the development of conditioned disgust reactions. Also, it provides the first demonstration of oxidative metabolic activity in brain regions involved in nausea-induced conditioned disgust. In addition, we found novel activation patterns of brain networks thought to be involved in the processing of flavors and their hedonic value. We highlighted an accumbens-amygdala interaction that may be involved in both appetitive and aversive reward, and a central role of the PBN nucleus in processing latent inhibition in conditioned taste aversion learning.

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References

- [1] Stockhorst, U., Steingrueber, H.J., Enck, P., Klosterhalfen, S., Pavlovian conditioning of nausea and vomiting, *Auton. Neurosci-Basic*. 129 (2006) 50-57.
- [2] Molassiotis, A., Managing nausea and vomiting after cancer treatments: patients still suffer unnecessarily, *Eur J Oncol Nurs*. 9 (2005) 4–5.
- [3] Roscoe, J.A., Morrow, G.R., Aapro, M.S., Molassiotis, A., Olve, I. Anticipatory nausea and vomiting. *Support Care Cancer*, 19 (2011) 1533-1538.
- [4] Stockhorst, U., Enck, P., Klosterhalfen, S., Role of classical conditioning in learning gastrointestinal symptoms, *World JGastroenterol*. 13 (2007) 3430–3437.
- [5] Limebeer, C.L., Hall, G., Parker, L.A., Exposure to a lithium-paired context elicits aping in rats: A model of anticipatory nausea, *Physiol. Behav*. 88 (2006) 398-403.
- [6] Rodríguez, M., López, M., Symonds, M., Hall, G., Lithium-induced context aversion in rats as a model of anticipatory nausea in humans, *Physiol. Behav.*, 71 (2000) 571-579.
- [7] Symonds, M., Hall, G., Contextual conditioning with an illness US is attenuated by the antiemetic ondansetron, *Psychobiology*, 28 (2000) 360-366.
- [8] Lubow, R. E., *Latent inhibition and conditioned attention theory*, Cambridge University Press, New York, 1989.
- [9] Lubow, R. E., Conditioned taste aversion and latent inhibition: A review, in: Reilly, S., Schachtman, T. R. (Eds.), *Conditioned taste aversion. Behavioral and neural processes*, NY: Oxford University Press, New York, 2009, pp. 58-73.
- [10] Grill, H.C., Norgren, R., The taste reactivity test: I. mimetic responses to gustatory stimuli in neurologically normal rats, *Brain Res*. 143 (1978) 263-279.
- [11] Parker, L.A., Rewarding drugs produce taste avoidance, but not taste aversion, *Neurosci. Biobehav. Rev*. 19 (1995) 143-151.

- [12] Parker, L.A., Taste avoidance and taste aversion: Evidence for two different processes, *Learn & Behav.* 31(2) (2003) 165-172.
- [13] Parker, L.A., Conditioned flavor avoidance and conditioned gaping: Rat models of conditioned nausea, *Eur J Pharmacol.* 722 (2014) 122-133.
- [14] López, M., Gasalla, P., Vega, M., Rock, E.M., Tuerke, K.S., Bedard, H., Parker, L.A., Latent inhibition of conditioned disgust reactions in rats, *Learn & Behav.* 38 (2010) 177-186.
- [15] Yamamoto, T., Yasoshima, Y., Electrophysiological representation of taste memory. *Neural plasticity and memory: From genes to brain imaging*, (2007) 113-128.
- [16] Yamamoto, T. A., Neural model for taste aversion learning, in: Kurihara, K., Suzuki, N., Ogawa, N., (Eds), *Olfaction and Taste XI*, Springer-Verlag, Tokyo: 1994, pp. 471.
- [17] Ashe, J. H., Nachman, M., Neural mechanisms in taste aversion learning, *Prog. Psychobiol. physiol. psychol.*, 9 (1980) 233-262.
- [18] Bernstein, I.L., Wilkins, E.E., Barot, S.K., Mapping conditioned taste aversion associations through patterns of c-fos expression, in: Reilly, S., Schachtman, T. R. (Eds.), *Conditioned taste aversion. Behavioral and neural processes*, NY: Oxford University Press, New York, 2009, pp. 58-73.
- [19] Núñez-Jaramillo, L., Ramírez-Lugo, L., Herrera-Morales, W., Miranda, M.I., Taste memory formation: latest advances and challenges, *Behav. Brain. Res.* 207(2) (2010) 232-48.
- [20] Yamamoto, T., Ueji, K., Brain mechanisms of flavor learning, *Front Syst Neurosci.*, 5 (2011) 1-7.
- [21] Garcia, J., Koelling, R.A., Relation of cue to consequence in avoidance learning, *Psychon. Sci.*, 4 (1966) 123-124.

- [22] Bures, J., Bermúdez-Rattoni, F., Yamamoto, T., Conditioned taste aversion. Memory of a special kind, Oxford University Press, Oxford, 1998.
- [23] Wong-Riley, M.T., Cytochrome oxidase: an endogenous metabolic marker for neuronal activity, Trends Neurosci. 12(3) (1989) 94-101.
- [24] Gonzalez-Lima, F., Jones, D., Quantitative mapping of cytochrome oxidase activity in the central auditory system of the gerbil: a study with calibrated activity standards and metal intensified histochemistry, Brain Res. 660 (1994) 34–49.
- [25] Arias, N., Morán, J., Conejo, N., Arias, J. Sexual metabolic differences in the rat limbic brain, Psicothema, 25 (2013) 461-467.
- [26] Bruchey, A.K., Gonzalez-Lima, F., Enhanced metabolic capacity of the frontalcerebral cortex after Pavlovian conditioning, Neurosci. 152(2) (2008) 299–307.
- [27] Conejo, N.M., González-Pardo, H., Gonzalez-Lima, F., Arias, J.L., Spatial learning of the water maze: progression of brain circuits mapped with cytochrome oxidase histochemistry, Neurobiol. Learn. Mem. 93(3) (2010) 362–371.
- [28] Conejo, N.M., Cimadevilla, J.M., González-Pardo, H., Méndez-Couz, M., Arias, J.L., Hippocampal inactivation with TTX impairs long-term spatial memory retrieval and modifies brain metabolic activity, Plos One, 8(5) (2013) e64749.
- [29] Fidalgo, C., Conejo, N.M., Gonzalez-Pardo, H., Lazo, P.S., Arias, J.L., A role for dorsal and ventral hippocampus in response learning, Neurosci. Res. 73 (3) (2012) 218-23.
- [30] Gonzalez-Lima, F., A. Cada, A., Quantitative histochemistry of cyto-chrome oxidase activity: theory, methods, and regional brain vulnerability, in: F. Gonzalez-Lima (Ed.), Cytochrome Oxidase in Neuronal Metabolism and Alzheimer’s Disease, Plenum, New York, London, 1998, pp. 55–90.

- [31] Méndez-Couzo, M., Conejo N.M., Vallejo, G., Arias, J.L., Brain functional network changes following Prelimbic area inactivation in a spatial memory extinction task, *Behav Brain Res.* 287 (2015) 247-55.
- [32] Sampedro-Piquero, P., Zancada-Menendez, C., Begega, A., Mendez, M., Arias, J.L., Effects of forced exercise on spatial memory and cytochrome c oxidase activity in aged rats, *Brain Res.* 1502 (2013) 20–29.
- [33] Sakata, J.T., Coomber, P., Gonzalez-Lima, F., Crews, D., Functional connectivity among Limbic brain areas: differential effects of incubation temperature and gonadal sex in the leopard gecko, *Eublepharis macularis*, *Brain. Behav. Evol.* 55 (2009) 139-151.
- [34] Grill, H. C., Part II: Physiological substrates of conditioned food aversions: Introduction: Physiological mechanism in conditioned taste aversions, *ANN NY ACAD SCI.* 443 (1998) 67-88.
- [35] Paxinos G, Watson G., *The Rat Brain in Stereotaxic Coordinates.* (Compact Third Edition), Academic Press, New York, 1997.
- [36] Shao, J., Dongsheng, T., *The Jackknife and bootstrap*, New York: Springer-Verlag, 1995.
- [37] Dwyer, D.M., Gasalla, P., López, M., Non reinforced flavor exposure attenuates the effects of conditioned taste aversion on both flavor consumption and cue palatability, *Learn & Behav.* 41 (2013) 390-401.
- [38] Ferreira, G., Ferry, B., Meurisse, M., Levy, F., Forebrain structures specifically activated by conditioned taste aversion, *Behav. Neurosci.* 120 (2006) 952-62.
- [39] Hernádi, I., Karádi, Z., Vígh, J., Petykó, Z., Egyed, R., Berta, b., Lénárd, L., Alterations of conditioned taste aversion after microiontophoretically applied neurotoxins in the medial prefrontal cortex of the rat, *Brain Res Bull.* 53 (2000) 751-758.

- [40] Karádi, Z., Lukáts, B., Papp, Sz., Egyed, R., Lénárd, L., Takács, G., Involvement of forebrain glucose-monitoring neurons in taste information processing electrophysiological and behavioral studies, *Chem Senses*. 30 (2005) 168-169.
- [41] Yamamoto, T., Neural substrates for the processing of cognitive and affective aspects of taste in the brain, *Arch. Histol. Cytol.* 69(4) (2006) 243-255.
- [42] Schafe, G.E., Bernstein, I.L., Forebrain contributions to the induction of a brainstem correlate of conditioned taste aversion. 1. The amygdala, *Brain Res.* 741 (1996) 109-16.
- [43] Ledoux, J.E., Emotion circuits in the brain, *Annu. Rev. Neurosci.* 23 (2000) 155-84.
- [44] Kosobud, A.E., Harris, G.C., Chapin, J.K., Behavioral associations of neuronal activity in the ventral tegmental area of the rat. *J. Neurosci.* 14 (1994) 7117–7129.
- [45] Park, T.H., Carr, K.D., Neuroanatomical patterns of fos-like immunoreactivity induced by a palatable meal and meal-paired environment in saline-and naltrexone-treated rats, *Brain Res.* 805 (1998) 169–180.
- [46] Shimura, T., Kamada, Y., Yamamoto, T., Ventral tegmental lesion reduce overconsumption of normally preferred taste fluid in rats, *Behav. Brain Res.* 134 (2002) 123-130.
- [47] Norgren, R., Taste pathways to hypothalamus and amygdala, *J. Comp. Neurol.* 166 (1976) 17–30.
- [48] Oades, R.D., Halliday, G.M., Ventral tegmental (A10) system: neurobiology: 1. Anatomy and connectivity, *Brain Res.* 434 (1987) 117–165.
- [49] Hajnal, A., Norgren, R., Taste pathways that mediate accumbens dopamine release by sapid sucrose, *Physiol Behav.* 84 (2005) 363-369.
- [50] Berridge, K.C., Robinson, T.E., What is the role of dopamine in reward: hedonic impact, reward learning, or incentive salience?, *Brain Res.* 28 (1998) 309-369.

- [51] Berridge, K.C., Liking and Wanting food rewards: brain substrates and roles in eating disorders, *Physiol. Behav.* 97 (5) (2009) 537-50.
- [52] Mark, G.P., Blander, D.S., Hoebel, B.G., A conditioned stimulus decreases extracellular dopamine in the nucleus accumbens after the development of learned taste aversion, *Brain Res.* 551 (1991)308-10.
- [53] Kho, M.T., Wilkins, E.E., Bernstein, I-L., Novel tastes elevate c-fos expression in the central amygdala and insular cortex: implication for taste aversion learning, *Behav Neurosci.* 117 (2003) 1416-1422.
- [54] Nachman, M., Ashe, J.H., Effects of basolateral amygdala lesions on neophobia, learned taste aversion, and sodium appetite in rats, *J. Comp. Physiol. Psychol.* 87 (1974) 622-643.
- [55] Reilly, S., Bornovalova, M.A., Conditioned taste aversion and amygdala lesions in the rat: a critical review, *Neurosci. Biobehav. Rev.* 29 (2005) 1067-1080.
- [56] Bures, J., Buresova, O., Reversible lesions allow reinterpretation of system level studies of brain mechanism of behavior, *Concepts Neurosci.* 1 (1990) 69-89.
- [57] Reilly, S., The parabrachial nucleus and conditioned taste aversion, *Brain Res. Bull.* 48(3) (1999) 239-254.
- [58] Agüero, A., Arnedo, M., Gallo, M., Puerto, A., The functional relevance of the lateral parabrachial nucleus in lithium chloride-induced aversion learning, *Pharm. Biochem. Behav.* 45 (1993) 973-978.
- [59] Reilly, S., Grigson, P. S., Norgren, R., Parabrachial nucleus lesions and conditioned taste aversion: Evidence supporting an associative deficit, *Behav. Neurosci.* 107 (1993) 1005-1017.
- [60] Sakai, N., Yamamoto, T., Conditioned taste aversion and c-fos expression in the rat brainstem after administration of various USs, *NeuroReport.* 8 (1997) 2215-2220.

Figure legends

Figure 1. Microphotographs showing some examples of coronal CO stained sections and schematic representations of the selected brain regions. The relative optical density of each region was measured taking five non-overlapping readings per region, in five consecutive sections using a sampling window adjusted for each region's size (an example is shown for the PrL). As microphotographs show, CO has the advantage of being a method with a high-resolution outcome allowing more accurate and automated measurement of activity in specific brain regions.

Figure 2. Mean number of disgust reactions (top panel) and mean duration (in seconds) of appetitive reactions (bottom panel) elicited during the TR test for the different groups: preexposed (PE-CTA), non-preexposed (NPE-CTA), preexposed control (PE-CONT) and non-preexposed control (NPE-CONT). Error bars represent the standard error of mean (SEMs).

Figure 3. Schematic representations of interregional correlations of CO activity for the different groups. Solid lines represent high positive pairwise Pearson's correlations ($p < .05$), whereas negative correlations are represented by broken lines. Each quadrant represents a group correlation, namely, preexposed (PE-CTA), non-preexposed (NPE-CTA), preexposed control (PE-CONT) and nonpreexposed control (NPE-CONT). A network is defined when regions are linked by correlations and are independent link from other structures. Specifically, a simple network between PBN and VTA was found in all groups with the exception of NPE-CONT. In this group, results show two main networks, one between PBN and AMY, and other established by thalamus, mPFC and NAc. Additionally, PE-CONT

group show a more intricate network between mPFC, NAcb, AMY and thalamus. We highlight a NAcb-AMY network in PE-CTA group, as well as correlations within mPFC regions. Finally, a simple network between thalamus and NAcb was found in NPE-CTA group.

Figure(s)

FIGURE 1

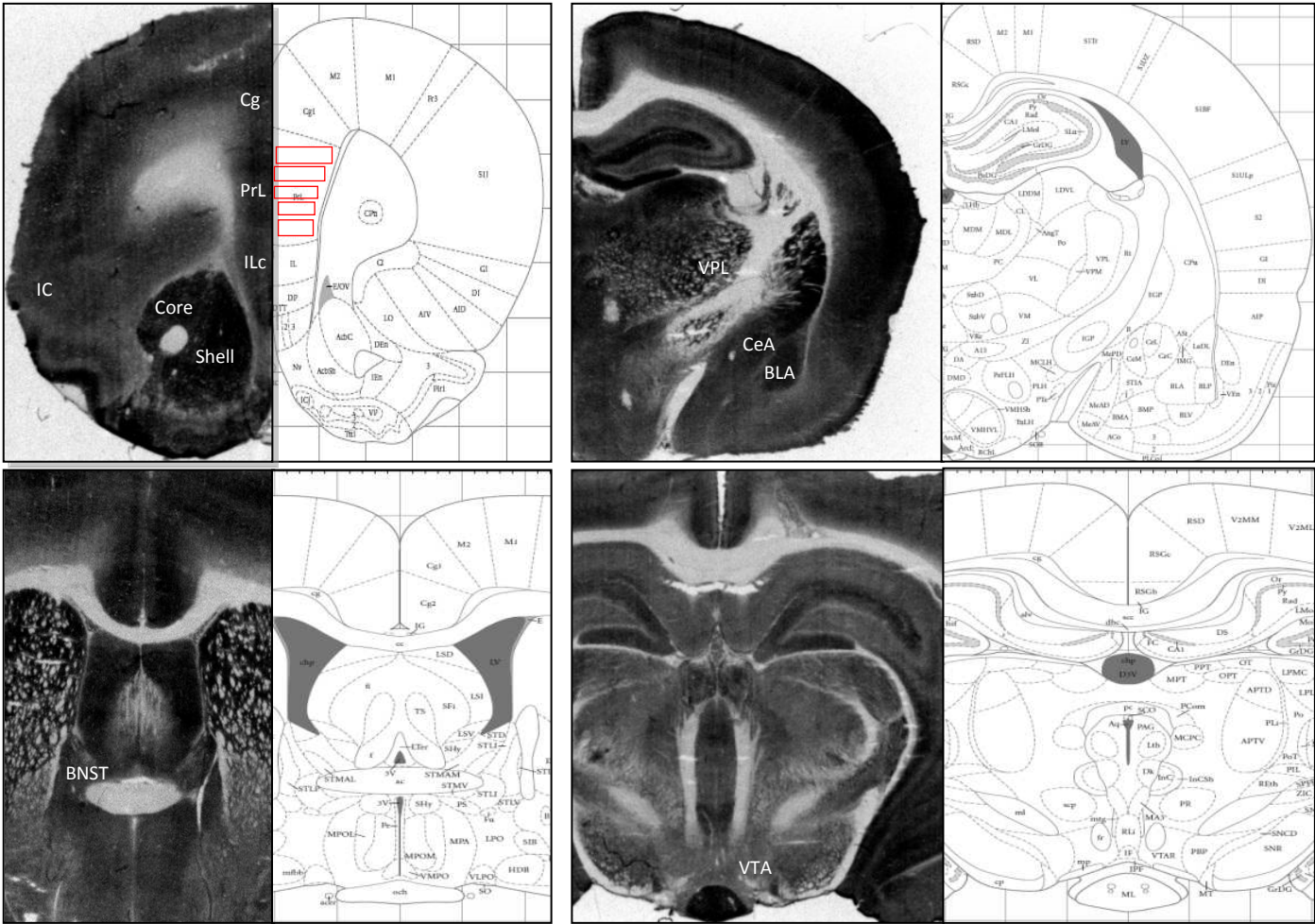


FIGURE 2

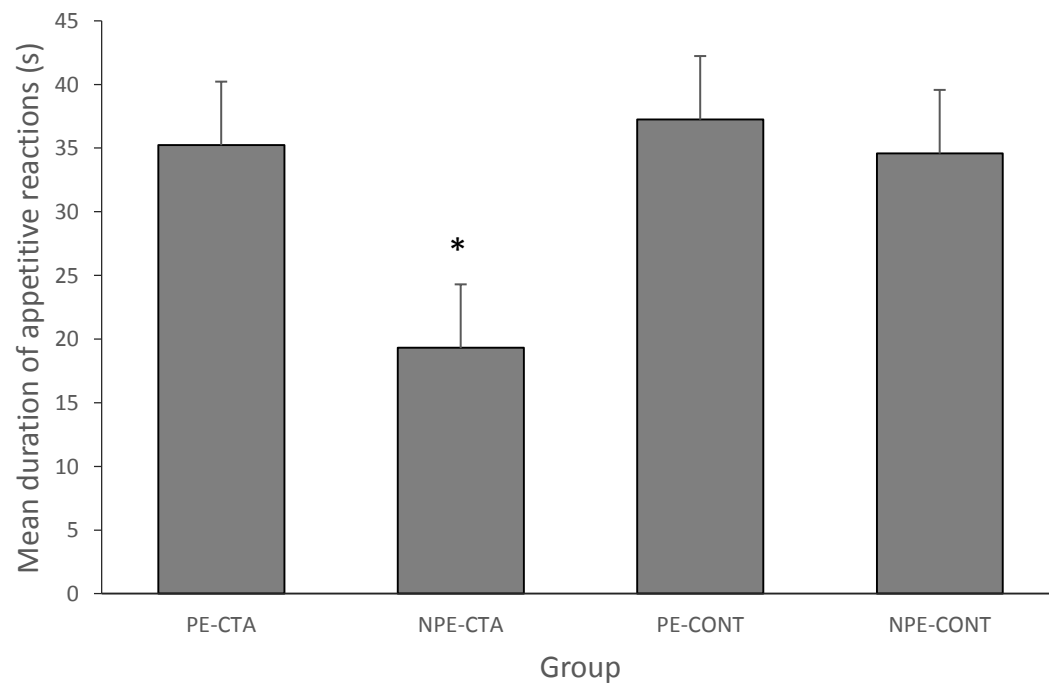
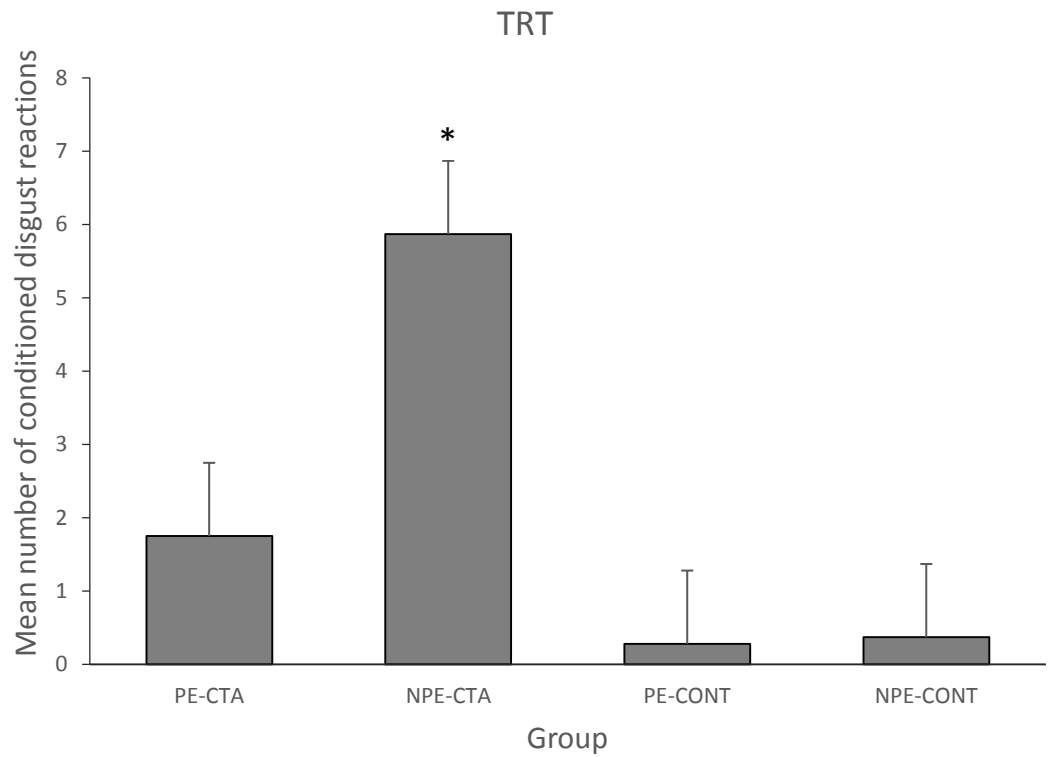


FIGURE 3

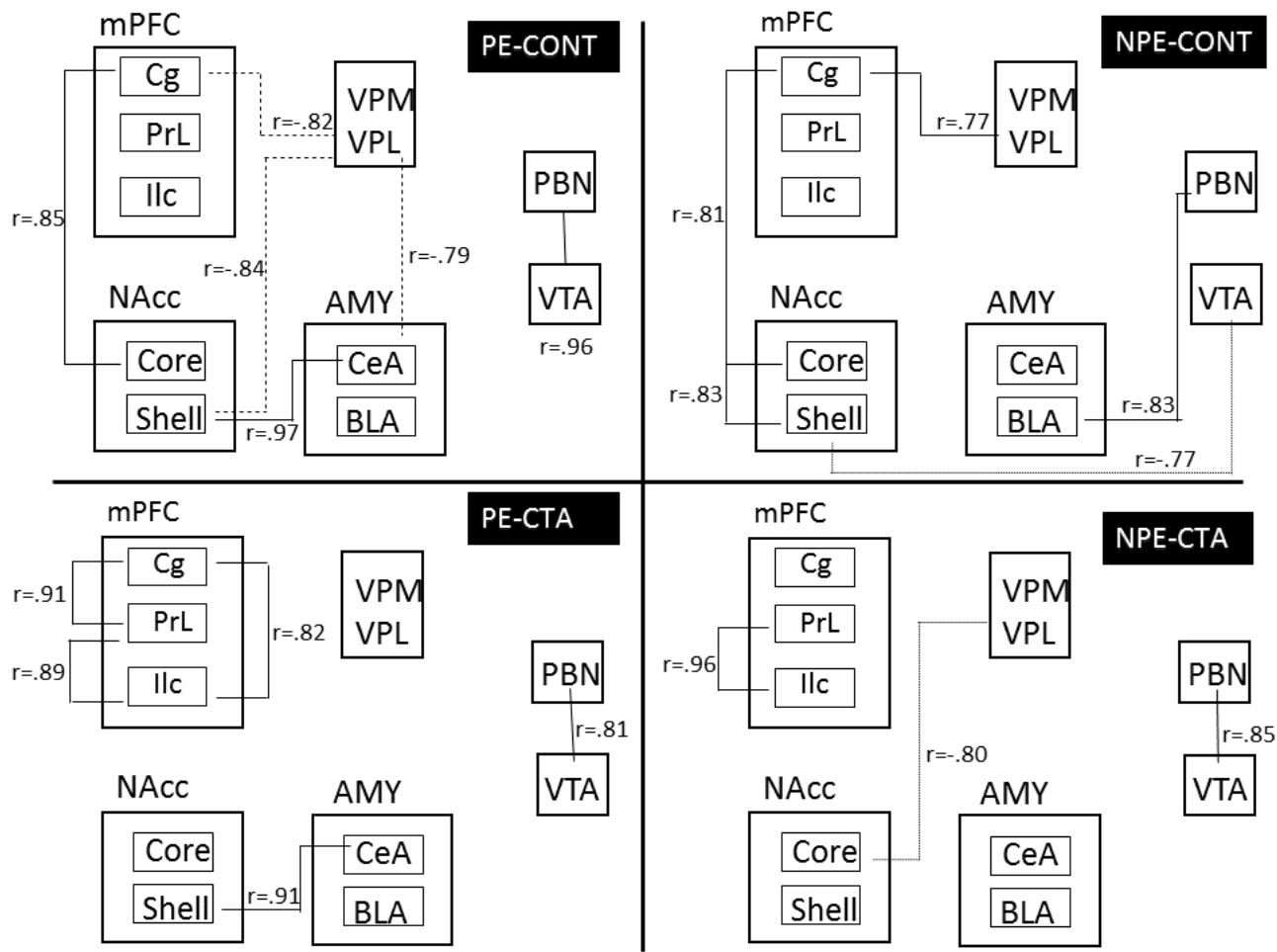


TABLE 1. Design of the experiment

Group	Preexposure	Conditioning	TR test
PE-CTA	4 x Sac (IO)	Sac (IO) → LiCl	Sac (IO)
NPE-CTA	4 x Water (IO)		
PE-CONT	4 x Sac (IO)	Sac (IO) / LiCl	
NPE-CONT	4 x Water (IO)		

Note: Taste aversion groups, PE-CTA (preexposed) and NPE-CTA (non-preexposed), were given a LiCl injection during conditioning of saccharin, and control groups, PE-CONT (preexposed) and NON-CONT (non-preexposed), received the LiCl injection on the day following the conditioning session. Sac = saccharin; IO = intraoral infusions; LiCl = lithium chloride injection.

TABLE 2. Cytochrome oxidase activity values of the measured regions

Regions	PE-CTA	NPE-CTA	PE-CONT	NPE-CONT	PE factor		CTA factor		PE *CTA Interaction	
					<i>F</i>	<i>p</i>	<i>F</i>	<i>p</i>	<i>F</i>	<i>p</i>
mPFC										
Cingulate	26.91 ± 1.28	27.51 ± 1.05	32.07 ± 0.80	33.13 ± 2.44	0.28	.603	11.74	.002	0.02	.883
Prelimbic	27.76 ± 1.84	27.81 ± 0.96	30.96 ± 0.50	30.19 ± 0.96	0.08	.768	5.29	.029	0.12	.737
Infralimbic	26.26 ± 1.6	26.72 ± 1.01	29.05 ± 0.63	28.99 ± 1.01	0.03	.866	4.59	.041	0.05	.831
Accumbens										
Core	34.38 ± 1.70	36.97 ± 1.82	34.76 ± 0.97	36.07 ± 3.00	0.89	.353	0.02	.897	0.09	.764
Shell	36.67 ± 1.71	37.36 ± 1.27	38.42 ± 1.08	41.52 ± 4.15	0.59	.451	1.43	.242	0.24	.630
IC	24.62 ± 1.56	27.76 ± 1.72	27.89 ± 0.56	26.29 ± 1.01	0.33	.571	0.45	.507	3.12	.089
BNET	25.75 ± 0.90	23.86 ± 1.00	28.26 ± 0.44	27.48 ± 0.89	2.40	.133	12.55	.001	0.41	.528
AMY										
CeA	29.76 ± 1.50	28.81 ± 1.16	29.84 ± 0.54	29.98 ± 2.18	0.07	.792	0.17	.684	0.13	.722
BLA	25.96 ± 2.41	27.57 ± 0.96	29.79 ± 0.57	31.13 ± 1.35	1.64	.212	4.29	.048	0.17	.683
VTA	12.61 ± 1.00	15.40 ± 1.00	17.82 ± 1.04	17.51 ± 1.28	1.28	.269	11.06	.003	1.99	.170
PBN	7.72 ± 0.79	9.16 ± 1.49	13.91 ± 1.63	11.82 ± 1.71	0.66	.425	4.07	.050	5.06	.033
Thalamic n.										
VPM	18.75 ± 1.79	21.99 ± 0.95	24.32 ± 0.95	21.09 ± 2.02	0.04	.848	5.08	.033	0.67	.420
VPL	28.71 ± 2.94	26.20 ± 2.83	21.10 ± 0.90	22.65 ± 2.34	0.60	.297	2.27	.143	4.35	.056

Note: Data represent Cytochrome oxidase values (mean ± SEM); 1 unit: 1 μmol of cytochrome c oxidase/min/g tissue wet

weight at 23° C. *F* values and *p* values are displayed for preexposure (PE) and conditioning (CTA) factors as well as for the

interaction between this two factors. A significant conditioning effect was found for the mPFC, BNET, BLA and VTA. Additionally,

a significant preexposure and conditioning interaction was found in the PBN. mPFC = Prefrontal cortex; IC = Insular Cortex; BNST

= bed nucleus of estria terminalis; AMY = amygdala (BLA = basolateral, CeA = central); VTA = ventral tegmental area; PBN=

parabrachial nucleus; VPM = ventral posteromedial thalamic nucleus; VPL = ventral posterolateral thalamic nucleus. PE-CTA =

preexposed group; NPE-CTA = non-preexposed group; PE-CONT = preexposed control group; NPE-CONT = non-preexposed

control group.